

## Introduction

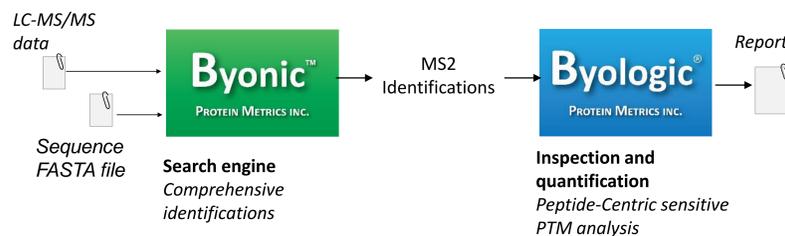
The pace of biosimilar development and pressures for rapid commercialization have led to the widespread adoption of mass spectrometry to support or disprove similarity to an innovator product. Analysts routinely use a wide array of biophysical and biochemical methods to characterize protein samples. As mass spectrometry development has progressed, the ability to have “multi-attribute methods” to efficiently report the quality attributes of a well-characterized biopharmaceutical product has become more realistic. It is only possible, however, with deep data mining of peptide maps and other data. In order to maximize the amount of information available and report it automatically, human intervention is at a premium. Although the development of mass spectrometer based methods are rapid, the tools needed to process the explosion of data are in short supply. Total and efficient analysis with the Protein Metrics suite of software reduces the human burden.

## Methods

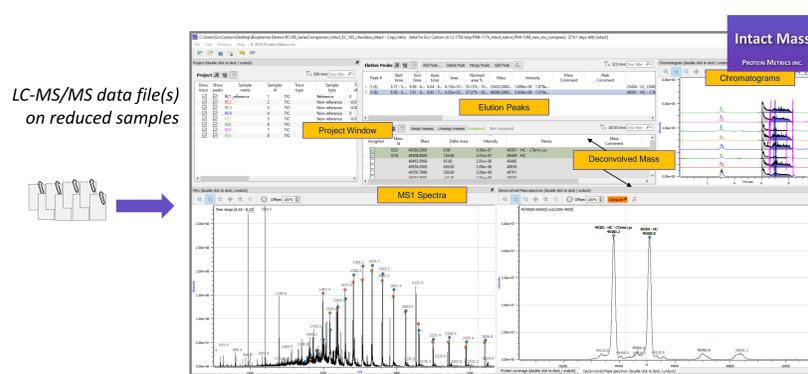
Four commercial lots of Remicade (Janssen) and Remsima (Celltrion) were sourced from the University of Michigan Hospital Pharmacy and South Korea, respectively. Samples were denatured, reduced, alkylated and digested with Promega’s low pH trypsin kit (Promega, CAS # CS1895A01). Digested samples were analyzed with an Orbitrap Q Exactive mass spectrometer (Thermo Fisher).

Raw MS/MS data were analyzed with Byonic™, Byologic® and , Intact Mass™, software packages (Protein Metrics). Samples were quantitatively compared with pivot summary reports specifically formatted for MAM.

### Data analysis workflow:



## Data Analysis & Reporting – Intact & Sub-unit Level



Peak #	Name	Remicade				Remsima			
		1	2	3	4	1	2	3	4
3	49381 - HC - C-term Lys	50.3	60.1	63	61.6	73.1	70.3	67.7	68.4
	49509 - HC	49.7	39.9	37	38.4	26.9	29.7	32.3	31.6

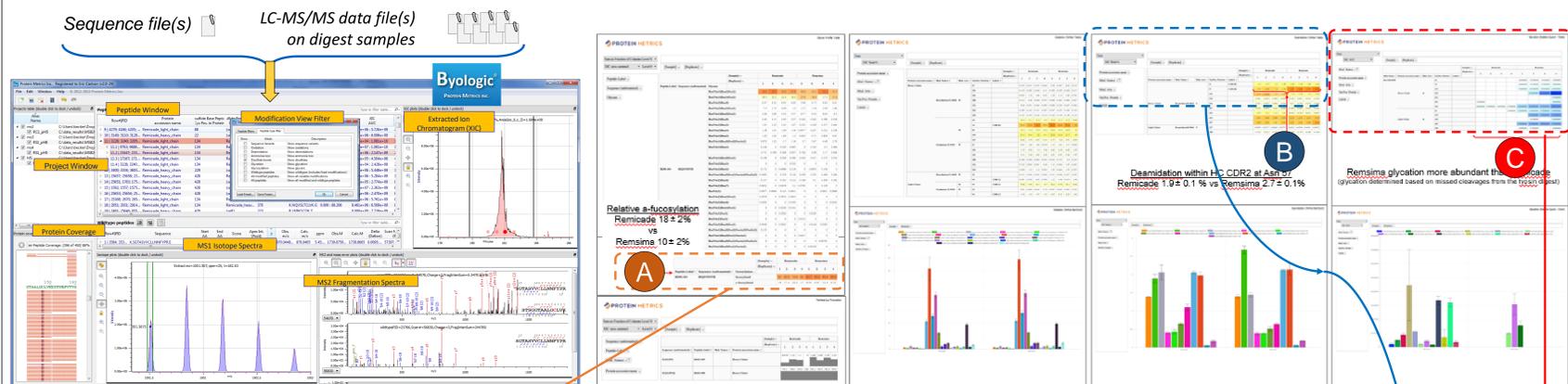
Observed increase in C-term Lys truncation in Remsima

Remicade 59±6%

vs

Remsima 70±2%

## Data Analysis & Reporting – Peptide Level



### Observed Differences

Peptide-Label	Sequence (unformatted)	Fucosylation	Remicade				Remsima			
			1	2	3	4	1	2	3	4
H296-304	EEQYNSTYR	fucosylated	81	82.6	79.8	83	91.7	90.3	86.6	90.5
		a-fucosylated	19	17.4	20.2	17	8.34	9.67	13.4	9.52

Relative a-fucosylation  
Remicade 18±2% vs Remsima 10±2%

Deamidation within HC CDR2 at Asn 57  
Remicade 1.9±0.1% vs Remsima 2.7±0.1%

Remsima glycation more abundant than Remicade (glycation determined based on missed cleavages from the trypsin digest)

Protein accession name	Mod. Names	Mod. AAs	Var.Pos.	Protein	Labels	Deamidation (%Mod Table)								
						Remicade				Remsima				
[Sample]	[Replicate]	[Sample]	[Replicate]	[Sample]	[Replicate]	1	2	3	4	1	2	3	4	
31	CDR H1		1.4	1.37	1.4	1.38	1.48	1.39	1.38	1.36				
57	CDR H2		1.79	1.98	2	1.79	2.7	2.68	2.57	2.82				

Mod. Names	Protein accession name	Mod. AAs	Var.Pos.	Protein	Labels	Glycation (Relative Quant - Table)							
						Remicade				Remsima			
[Sample]	[Replicate]	[Sample]	[Replicate]	[Sample]	[Replicate]	1	2	3	4	1	2	3	4
Heavy Chain	K		43			16900000	21200000	19500000	23900000				
			78			44400000	47100000	51700000	58900000				
			136			75400000	84700000	89700000	102000000				
			249					18100000	82500000	107000000			
			251							111000000			
			291		23400000								
			329		49400000	42900000	30200000	47900000	22600000	30200000	29200000	40900000	98900000
			49					16900000	12400000	13600000	17500000	19000000	
			149						31900000	31900000	31100000		
			169						32100000		11000000		
			183					27800000	33200000	34900000	31300000		
			190					26000000	37300000		44400000		

### Published Differences

Property	Remicade	Remsima
Sequence	Identical	Identical
TNF-α binding	Identical	Identical
FcγR-IIIa binding	Strong	Lower
ADCC assay	Strong	Lower
N-Glycan (Asn 300)	G0F, G1F, G2F, Man5	G0F, G1F, G2F
C-terminal lysine		More truncation

## Discussion and Conclusions

The commercial lots of Remicade (originator) and Remsima (biosimilar) show substantial similarity when compared for a list of variants, PTMs, and overall intact mass of major species.

This multi-attribute comparison does reveal some analytical differences between the samples. Using this multi-attribute method (MAM), we were able to efficiently highlight the differences in deamidation in the HC CDR2 region, relative glycation, C-terminal Lys truncation, and relative a-fucosylation of the glycan profile. (See notes A → D.)

As interest in MAM increases, it is essential that the historically separate strands of information are pulled together. Here, we present a method that copes not only with complex data from peptide mapping, which has been a focus of MAM, but also with the intact mass information, which provides identity and reports on major isoforms.

Reporting data, whether for MAM methods, or for process samples, or for assay development, relies on bringing together data sets from different sources and with varying skill sets. To scale this for consistent business processes, this requires a consistent platform, like the suite of software shown here. The Protein Metrics Biopharma Software Suite facilitates multi-attribute methods analysis with a platform able to compare across samples and produce reports that summarize PTM and peptide level variants and as well as Intact Mass reporting.

## Acknowledgment

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